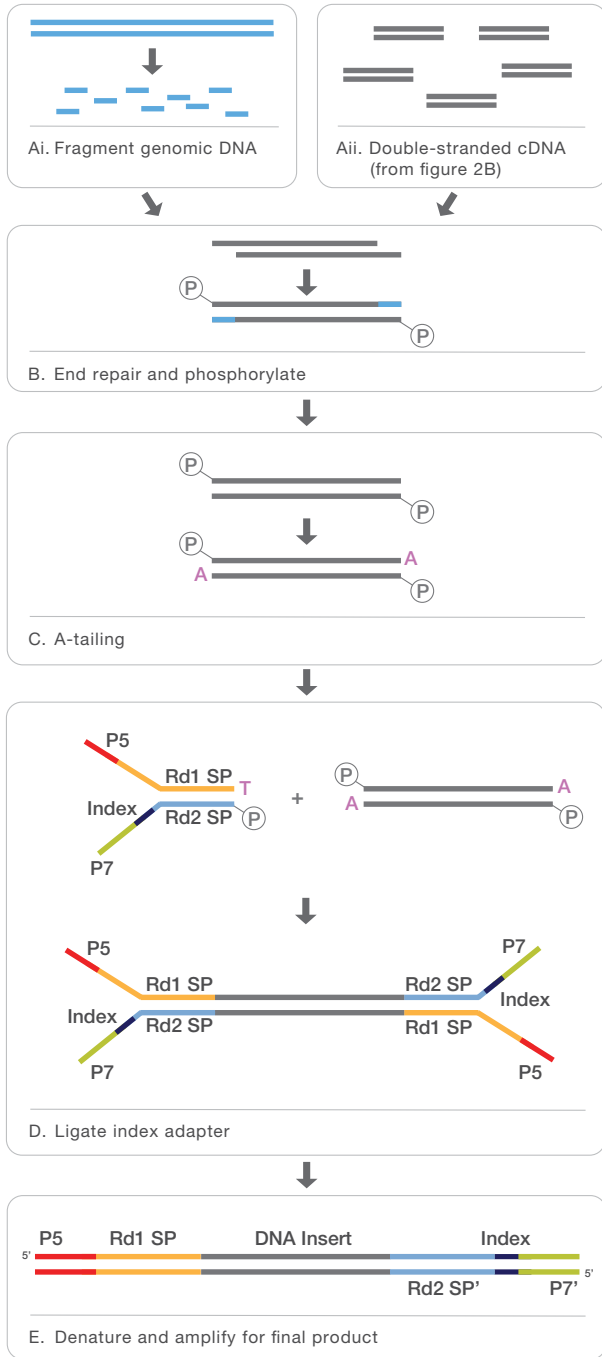


Figure 4: Adapter Ligation Results in Sequence-Ready Constructs without PCR



Library construction begins with either fragmented genomic DNA or double-stranded cDNA produced from total RNA (Figure 4A). Blunt-end fragments are created (Figure 4B) and an A-base is then added (Figure 4C) to prepare for indexed adapter ligation (Figure 4D). Final product is created (Figure 4E), which is ready for amplification on either the cBot or the Cluster Station.

to the A-tailed fragmented DNA. These newly redesigned adapters contain the full complement of sequencing primer hybridization sites for single, paired-end, and multiplexed reads. This eliminates the need for additional PCR steps to add the index tag and multiplex primer sites (Figure 4D). Following the denaturation and amplification steps (Figure 4E), libraries can be pooled with up to 12 samples per lane (96 sample per flow cell) for cluster generation on either cBot or the Cluster Station.

Master-mixed reagents and an optimized protocol improve the library construction workflow, significantly decreasing hands-on time and reducing the number of clean-up steps when processing samples for large-scale studies (Table 1). The simple and scalable workflow allows for high-throughput and automation-friendly solutions, as well as simultaneous manual processing for up to 96 samples. In addition, enhanced troubleshooting features are incorporated into each step of the workflow, with quality control sequences supported by Illumina RTA software.

Enhanced Quality Controls

Specific Quality Control (QC) sequences, consisting of double-stranded DNA fragments, are present in each enzymatic reaction of the TruSeq sample preparation protocol: end repair, A-tailing, and ligation. During analysis, the QC sequences are recognized by the RTA software (versions 1.8 and later) and isolated from the sample data. The presence of these controls indicates that its corresponding step was successful. If a step was unsuccessful, the control sequences will be substantially reduced. QC controls assist in comparison between experiments and greatly facilitate troubleshooting.

Designed for Automation

The TruSeq Sample Preparation Kits are compatible with high-throughput, automated processing workflows. Sample preparation can be performed in standard 96-well microplates with master-mixed reagent pipetting volumes optimized for liquid-handling robots. Barcodes on reagents and plates allow end-to-end sample tracking and ensure that the correct reagents are used for the correct protocol, mitigating potential tracking errors.

Part of an Integrated Sequencing Solution

Samples processed with the TruSeq Sample Preparation Kits can be amplified on either the cBot Automated Cluster Generation System or the Cluster Station and used with any of Illumina's next-generation sequencing instruments, including HiSeq™ 2000, HiSeq 1000, HiScan™SQ, Genome Analyzer_{IIx} (Figure 5).

Summary

Illumina's new TruSeq Sample Preparation Kits enable simplicity, convenience, and affordability for library preparation. Enhanced multiplexing with 24 unique indexes allows efficient high-throughput processing. The pre-configured reagents, streamlined workflow, and automation-friendly protocol save researchers time and effort in their next-generation sequencing pursuits, ultimately leading to faster discovery and publication.

Learn more about Illumina's next-generation sequencing solutions at www.illumina.com/sequencing.

