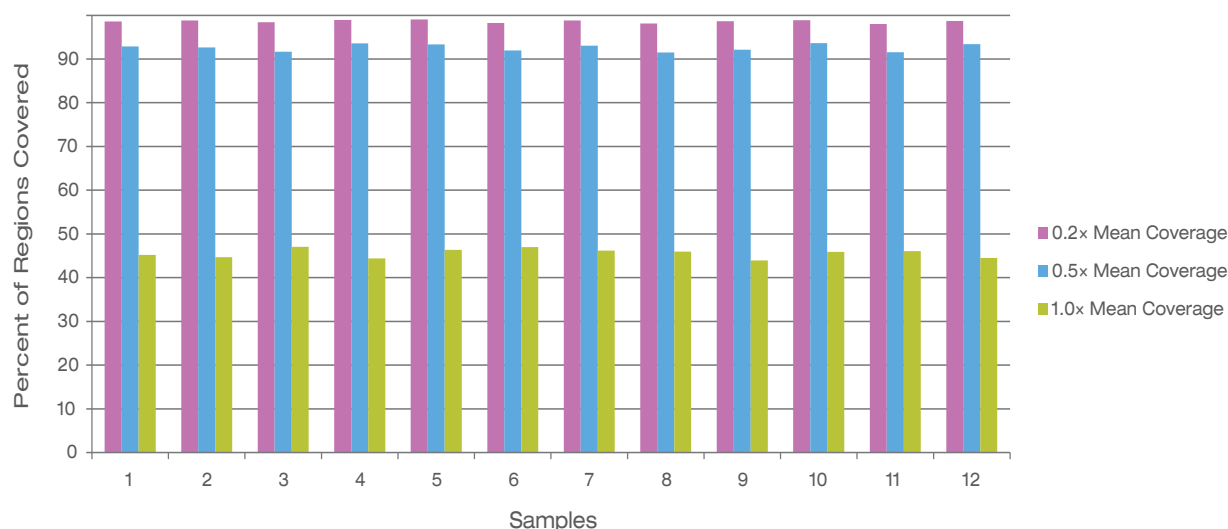


Figure 4: High Coverage Uniformity



Coverage uniformity is given for 12 samples with respect to the percentage of targeted regions at varying mean normalized read depths. The 12 samples were prepared and then simultaneously enriched using the TruSight Rapid Capture along with the TruSight Cancer Sequencing Panel. Pooled samples were sequenced across a MiSeq[®] standard flow cell, generating mean read depths of 130–230× (varying for each sample). Over 95% of bases (~250 Kb) were covered at 0.2× mean coverage.

friendly workflow. TruSight Rapid Capture kits supporting lower throughput options are also available, allowing labs to cost-effectively run samples immediately instead of waiting to batch.

The process starts with rapid Nextera-based sample prep to convert input genomic DNA into adapter-tagged libraries (Figure 3A). This rapid prep requires only 50 ng of input DNA and takes less than 3 hours for a plate of 96 samples. Nextera fragmentation of DNA simultaneously fragments and tags DNA without the need for mechanical shearing. Integrated sample barcodes then allow the pooling of up to 96 samples for a single Rapid Capture pull down. Next, libraries are denatured into single-stranded DNA (Figure 3B) and biotin-labeled probes specific to the targeted region are used for the Rapid Capture hybridization (Figure 3C). The pool is enriched for the desired regions by adding streptavidin beads that bind to the biotinylated probes (Figure 3D). Biotinylated DNA fragments bound to the streptavidin beads are magnetically pulled down from the solution (Figure 3E). The enriched DNA fragments are then eluted from the beads and hybridized for a second Rapid Capture. This entire process is completed in only 1.5 days, enabling a single researcher to efficiently process up to 288 samples at one time—all without automation.

Data Analysis

Sequence data generated from TruSight Cancer enriched libraries are analyzed by the on-instrument MiSeq Reporter (MSR) software. After demultiplexing and FASTQ file generation, the software uses the Burrows-Wheeler Aligner (BWA) to align the reads against the hg19 homo sapiens reference genome to create BAM files. The Genome Analysis Toolkit (GATK) is then used to perform variant analysis for the target regions specified in the manifest file. The output of GATK are VCF files, which are text files that contain SNPs, indels, and other structural variants.

High Data Quality

With TruSight Cancer and TruSight Rapid Capture, researchers can be confident in the quality of sequencing data generated from pooled multisample libraries. Each sample is sequenced with high coverage uniformity across the target region, with 95% of exons covered at a minimum coverage of 20× (Figure 4). This uniformity applies to smaller exons (< 150 bp) as well as long coding exons.

Summary

The TruSight Cancer Sequencing Panel enables researchers to access an expert-defined content set for analyzing variation within genes previously linked with a predisposition towards cancer. The optimized probe set provides comprehensive coverage of the targeted regions with high coverage uniformity for identifying a large number of variants. Combining this content with the TruSight Rapid Capture method enables a fast, easy workflow, requiring low sample DNA input, generating a highly efficient resequencing solution to accelerate detection of genes associated with cancer.

Learn More

To learn more about TruSight Cancer Sequencing Panel, TruSight Rapid Capture kits, and Illumina next-generation sequencing technology, visit www.illumina.com/trusightcancer.

Reference

1. Optimizing Coverage for Targeted Resequencing Technical Note (www.illumina.com/documents/products/technote/technote_optimizing_coverage_for_targeted_resequencing.pdf)

